

## Introduction

Biodiesel is a potential renewable, clean, and green energy. Using *Chlorella protothecoides* can help pave the path for this potential fuel. *C. protothecoides* has the ability to be grown both heterotrophically and autotrophically, and when grown heterotrophically growth restriction that burden many other strains are reduced. Optimization of growing *C. protothecoides* heterotrophically can greatly help us understand this species and the procedures to maximize lipid production genetically, greatly increase cell growth and density, and reduce lipid loss in the extraction process.

## Gene Alternation



**Figure 1:** *C. protothecoides* that underwent UV exposure and was diluted 1:100,000 solution. These were then grown up in a flask and retested for lipid content.

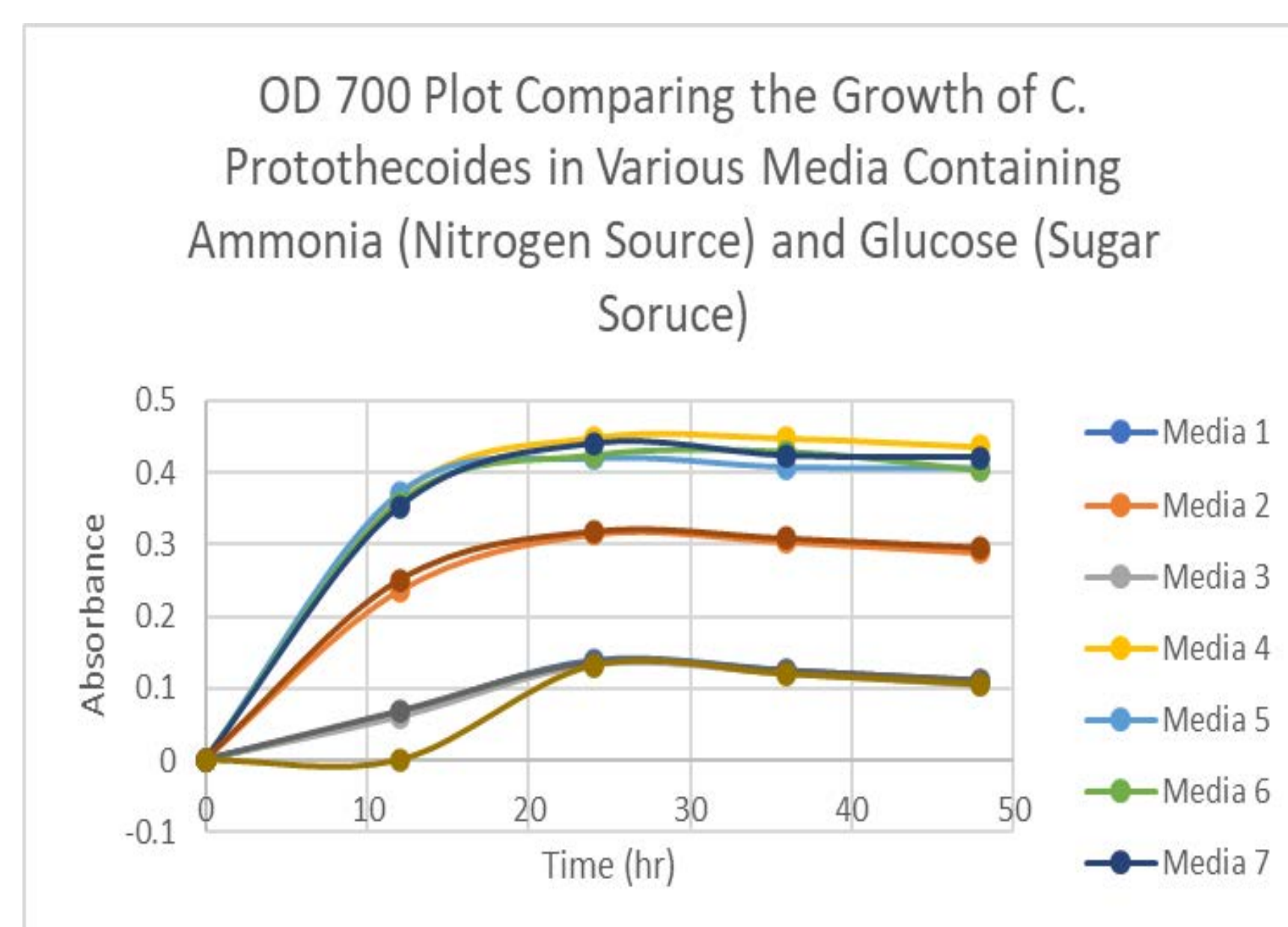
Lipid content was optimized by mutating *C. protothecoides* DNA by UV radiation. Cells surviving UV radiation were then treated with Nile Red stain and the lipid content observed under a fluorescent microscope. Sample were then regrown through to create a serial enrichment of lipid content

## Growth Optimization

Growth was optimized considering two main factors; ammonia and glucose. This was achieved by varying the volume of stock A, B, C and D, fully detailed in **Figure 2**, in the growing media. Samples were grown for 48 hours after which absorbances were collected. Results are shown in figure 3 with the greatest absorbance from stock B.

| Group A 10x Stock |             |                                    |
|-------------------|-------------|------------------------------------|
| 98.57             | grams/liter | MSG monohydrate                    |
| 6.86              | grams/liter | NaCl                               |
| 3.18              | grams/liter | CaCl2 monohydrate                  |
| 10.95             | grams/liter | KCl                                |
| 54.76             | grams/liter | MgSO4 septahydrate                 |
| 4.05              | grams/liter | Ammonium sulfate                   |
| 65.71             | grams/liter | Yeast extract                      |
| 5.70              | grams/liter | KH2PO4                             |
| 1.10              | grams/liter | Dow 1520US antifoam                |
| Group B 10x stock |             |                                    |
| 52.667            | grams/liter | FeSO4 septahydrate                 |
| 94.249            | grams/liter | citric acid                        |
| 15.844            | grams/liter | MnCl2 tetrahydrate                 |
| 15.844            | grams/liter | ZnSO4 septahydrate                 |
| 0.204             | grams/liter | CoCl2 hexahydrate                  |
| 0.204             | grams/liter | NaMoO4 dihydrate                   |
| 10.580            | grams/liter | CuSO4 pentahydrate                 |
| 10.580            | grams/liter | NiSO4 hexahydrate                  |
| Group C 10x stock |             |                                    |
| 11.2125           | grams/liter | thiamine-HCl                       |
| 0.184             | grams/liter | vitamin B12                        |
| 3.835             | grams/liter | pantothenic acid hemi-calcium salt |
| Group D 10x Stock |             |                                    |
| 300               | grams/liter | glucose                            |

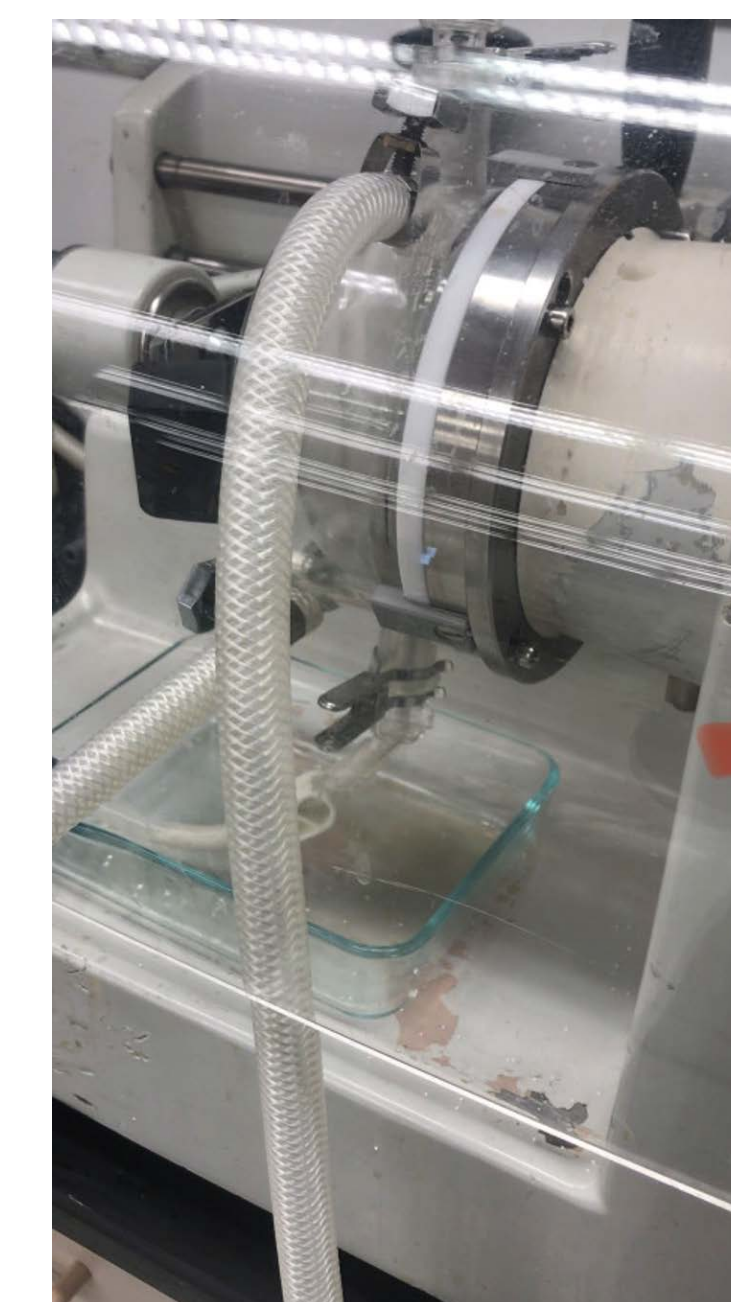
**Figure 2:** Stock media recipes used in varying amount



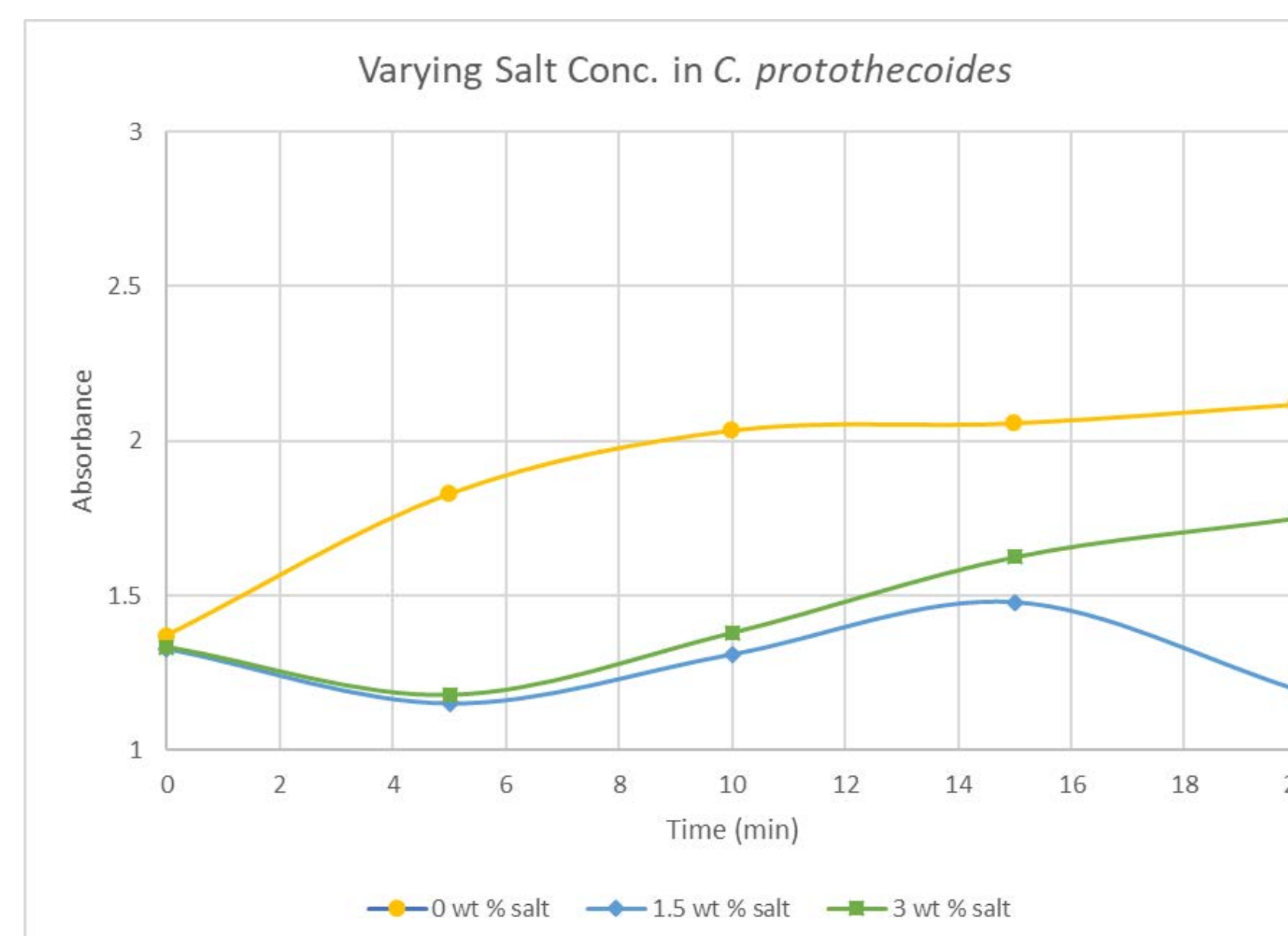
**Figure 3:** Absorbance of varying samples over time.

## Extraction

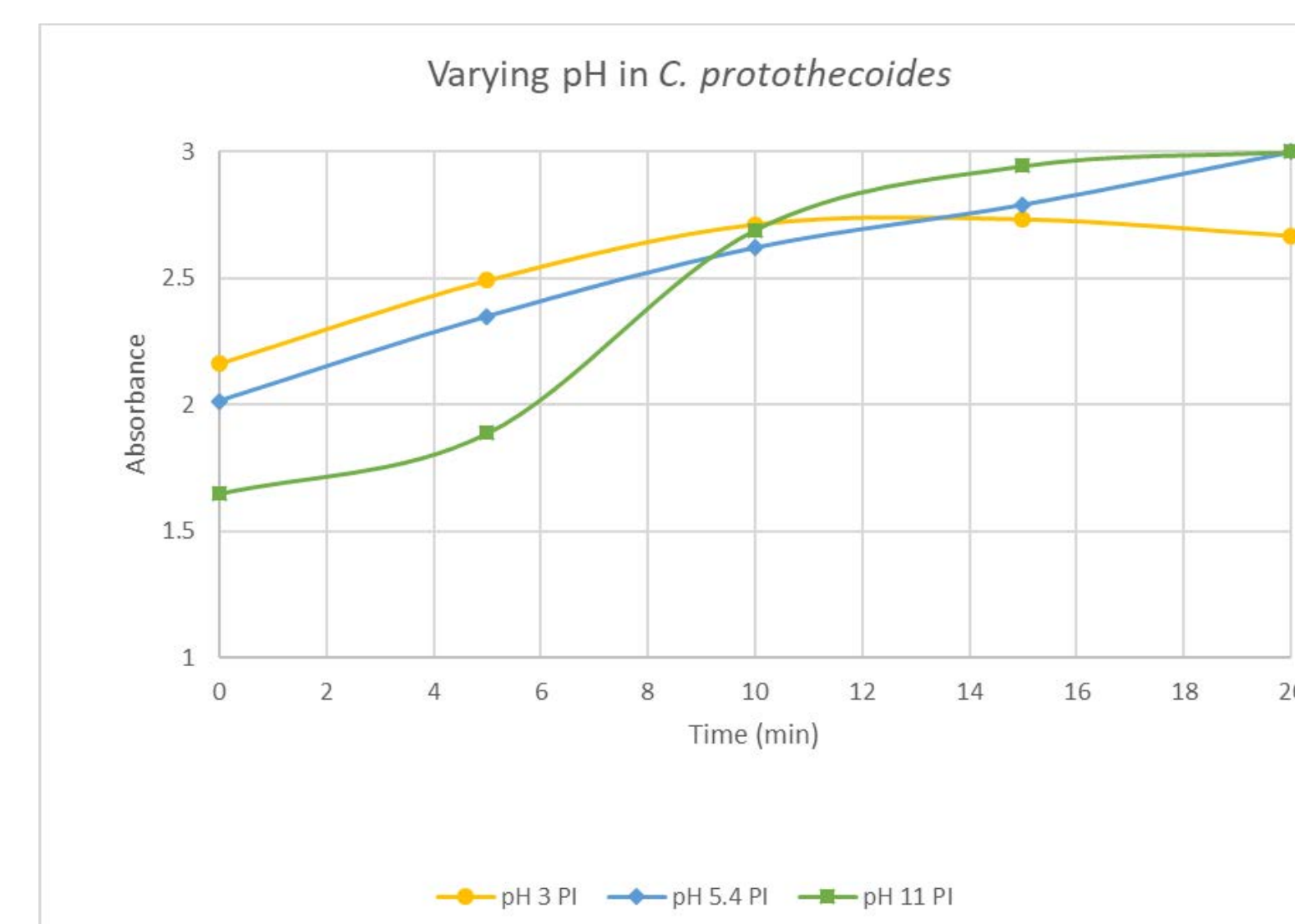
Extraction optimization was achieved by varying salt, pH, and milling time. Samples were collected every five minutes for absorbance testing using propidium iodide and spectrophotometry. Ten minutes with no added salt and original pH resulted in the best extraction.



**Figure 4:** 300 mL glass bead mill



**Figure 5:** Absorbance of varying samples over time in the bead mill with wt% salt adjusted using sodium chloride.



**Figure 6:** Absorbance of varying samples over time in the bead mill with pH adjusted using sulfuric acid and sodium hydroxide.

## Conclusions

The team has explored several stages of optimizing *Chlorella protothecoides* for the production of biodiesel. The current data results suggest using a growing media in comparison to stock B for the most dense growth of *Chlorella protothecoides* followed by ten minutes in the bead mill using 0.8-1  $\mu\text{m}$  beads. No adjustment to cell culture are needed for the best milling results. Future studies will provide for better optimization and reduced material input.

## Future Work

The following are suggestions to further improve biodiesel from *C. protothecoides*.

- Kill samples need to be carried onward to test for lipid percentage.
- Media variants such as glycerol-ammonia, glucose-urea, and glycerol-urea experiments
- Will provide carbon and nitrogen sources for the algae while utilizing biofuel biproducts.
- Factors such as temperature and enzyme cocktails should considered in the milling process.

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